

Research paper**Forage yield and nutritive value of *Arachis* spp. genotypes in the Brazilian savanna***Producción de forraje y valor nutritivo de genotipos de *Arachis* spp. en el Cerrado brasileño*FRANCISCO D. FERNANDES¹, ALLAN K.B. RAMOS¹, MARCELO A. CARVALHO¹, GIOVANA A. MACIEL¹, GISELLE M.L. DE ASSIS² AND GUSTAVO J. BRAGA¹¹Empresa Brasileira de Pesquisa Agropecuária, Embrapa Cerrados, Planaltina, DF, Brazil. www.embrapa.br/cerrados²Empresa Brasileira de Pesquisa Agropecuária, Embrapa Acre, Rio Branco, AC, Brazil. www.embrapa.br/acre**Abstract**

Forage yield, nutritive value, ground cover and mineral concentration of 10 genotypes of *Arachis* spp. were evaluated over 3 years in Planaltina, Federal District, Brazil. Experimental plots were arranged in a randomized complete block design with 4 replications. Treatments were 5 genotypes of *A. pintoi* (accessions 2, 4, 5, 6 and 8), 2 genotypes of *A. repens* (accessions 1 and 7), 1 hybrid *A. pintoi* × *A. repens* (accession 3) and 2 *A. pintoi* cultivars, BRS Mandobi and Belmonte. All genotypes established well and achieved good ground cover in the first year. Cultivar Belmonte and accessions 2 and 4 produced the highest DM yields (means of 8.8, 8.5 and 8.8 t DM/ha/yr, respectively) throughout, while cv. BRS Mandobi and accession 6 were the worst (5.7 and 5.6 t DM/ha/yr). Most genotypes maintained ground cover above 80% throughout the study but cv. BRS Mandobi plus accessions 6 and 8 had declined to 60% or less by the third year. Mean crude protein concentration overall was 166 g/kg with a range of 154–182 g/kg among genotypes. There was no major genotypic variation in mineral concentrations, which in all cases were considered marginal to adequate for tropical forage legumes. In conclusion, accessions 2 and 4 (BRA-039799 and BRA-039187, respectively) of *A. pintoi* are considered the most promising forage peanut options under the edaphoclimatic conditions of the experimental site. More effort is needed to improve seed set in these genotypes to increase adoption by farmers. Their persistence under grazing and impact on production should also be demonstrated.

Keywords: *Arachis pintoi*, *Arachis repens*, crude protein, digestibility, minerals.**Resumen**

En Planaltina, Distrito Federal, Brasil, se evaluaron la producción de forraje, el valor nutritivo, la cobertura del suelo y la concentración mineral de 10 genotipos de *Arachis* spp. durante 3 años. Las parcelas experimentales se dispusieron en un diseño de bloques completos al azar con 4 repeticiones. Los tratamientos consistieron en 5 genotipos de *A. pintoi* (accesiones 2, 4, 5, 6 y 8), 2 genotipos de *A. repens* (accesiones 1 y 7), 1 híbrido *A. pintoi* × *A. repens* (accesión 3) y 2 cultivares de *A. pintoi*, BRS Mandobi y Belmonte. Todos los genotipos presentaron buen establecimiento y buena cobertura del suelo en el primer año. Considerando los 3 años del experimento, el cultivar (cv.) Belmonte y las accesiones 2 y 4 presentaron los rendimientos de materia seca (MS) más altos, con promedios de 8.8, 8.5 y 8.8 t MS/ha por año, respectivamente, mientras que cv. BRS Mandobi y la accesión 6 fueron los menos productivos, con promedios de 5.7 y 5.6 t MS/ha por año, respectivamente. La mayoría de los genotipos mostraron una cobertura del suelo superior a 80% durante la duración del estudio; no obstante el cv. BRS Mandobi y las accesiones 6 y 8 presentaron una reducción al 50–60% al finalizar el tercer año. En total, la concentración promedio de proteína cruda fue de 166 g/kg con un rango de 154–182 g/kg entre genotipos. Las concentraciones de minerales no mostraron mayores variaciones entre genotipos y se consideraron marginales a adecuadas para leguminosas forrajeras tropicales. En conclusión, las accesiones 2 y 4 de

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A. pintoii (BRA-039799 y BRA-039187, respectivamente) mostraron ser las opciones más prometedoras bajo las condiciones edafoclimáticas del sitio experimental. Se requieren esfuerzos para mejorar la producción de semilla de estos materiales para lograr una mayor adopción por los productores. Además su persistencia bajo pastoreo y su impacto en la producción deben demostrarse.

Palabras clave: *Arachis pintoii*, *Arachis repens*, digestibilidad, minerales, proteína cruda.

Introduction

Cattle farms in the Brazilian savannas (Cerrados) are practically totally dependent on pastures sown with African grasses, which may be exposed to overgrazing, low soil fertility and water deficits (Oliveira et al. 2004). Despite being considered a sustainable way to improve livestock productivity at low cost (Boddey et al. 1997), the sowing of legumes by farmers is not a regular practice in Brazil. The use of tropical forage legumes is restricted to particular areas, like the northwestern humid tropics in Acre State, where *Pueraria phaseoloides* and *Arachis pintoii* are sown in mixed grass-legume pastures (Assis et al. 2013). Limited numbers of suitable legume cultivars is one of the barriers to the use of legumes in forage-based production systems. Furthermore, low persistence of legumes, cost of seed, the need for vegetative propagation in some cases and undefined grazing management recommendations also contribute to their limited usage (Wünscher et al. 2004), as opposed to the wide-spread use of more resilient and accessible grasses like *Brachiaria* spp.

Several *Arachis* species with forage potential (“forage peanut”) are endemic in Brazil (Valls and Simpson 1994), presenting satisfactory performance, especially when sown in humid areas (>1,500 mm rainfall per year). Within the *Arachis* genus, *A. pintoii* is the most commonly sown for forage in the world and the only one with registered cultivars in Brazil. Approximately 0.14 M ha are cultivated with cv. Belmonte (*A. pintoii*) in the State of Acre, but expansion of this area is slow owing to the predominant use of vegetative propagation (Assis et al. 2013). The Australian cv. Amarillo (*A. pintoii*) is the only cultivar for which seed is available in the Brazilian market, and it is usually imported from Bolivia and Peru. Under savanna conditions, annual dry matter (DM) yields from *A. pintoii* of up to 13 t/ha have been reported, and for *A. repens* yields up to 6 t DM/ha have been obtained in a lowland area (Pizarro and Rincón 1994). In addition to tolerance to heavy grazing and adaptation to infertile soils (Rao and Kerridge 1994), *Arachis* presents high nutritive value and acceptability (Argel and Pizarro 1992) and supports better cattle performance than grass monocultures (Lascano 1994). However, because of irregular rainfall distribution in savannas, yield and persistence of

A. pintoii are not always satisfactory, especially when it is grown in highland areas. The objective of this study was to evaluate forage yield, ground cover, nutritive value and macronutrient concentrations of genotypes, cultivars and a hybrid of *Arachis* spp. in a Brazilian savanna (“Cerrado”) environment.

Materials and Methods

Experimental field and environmental conditions

The experiment was carried out in Planaltina, Federal District, Brazil (15°35' S, 47°42' W; 993 masl) during 3 consecutive years. The climate at the site is Aw according to Köppen-Geiger classification (Peel et al. 2007). Monthly rainfall and mean air temperature were recorded at 600 m from the experimental site and are presented along with the historical averages for annual rainfall and temperature (Table 1). The soil in the area is classified as an Oxisol, with very clayey texture, pH (in H₂O) 5.2, organic matter content of 0.22 g/kg, phosphorus (P) 6.6 mg/dm³ (Mehlich-I), potassium (K) 38.7 mg/dm³, calcium (Ca) 2.6 mg/dm³, magnesium (Mg) 0.64 mg/dm³ and aluminum (Al) 0.14 mg/dm³ in the 0–0.2 m soil layer. On 31 March 2008, after the first harvest of the experimental period, 25 kg K/ha as potassium chloride and 4 kg P/ha as single superphosphate were applied; this was the only fertilization during the entire evaluation period.

Arachis spp. germplasm sources

The treatments were 10 genotypes of *Arachis* spp. (Table 2), consisting of 5 genotypes of *Arachis pintoii* Krapov. & W.C. Greg. (accessions 2, 4, 5, 6 and 8), 2 genotypes of *Arachis repens* Handro (accessions 1 and 7), 1 inter-specific hybrid (accession 3) from a cross between *A. pintoii* cv. Amarillo and *A. repens* BRA-029220 and 2 cultivars of *A. pintoii* (cvv. Belmonte and BRS Mandobi). The genotypes and hybrid were drawn from the *Arachis* spp. germplasm collection of Embrapa Acre, located in Rio Branco, Acre State, Brazil. Cultivar Belmonte was collected in Bahia State, Brazil, and released in 1999 (Paganella and Valls 2002). Cultivar BRS Mandobi was obtained from a network of mass selection promoted by Embrapa in Brazil including the states Acre, Bahia and the Federal District (Assis et al. 2013).

Table 1. Monthly rainfall and mean temperature during the experimental period and historical means (1974–2006) for Planaltina, Federal District, Brazil.

Month	Rainfall (mm)					Temperature (°C)				
	2007	2008	2009	2010	Avg. ¹	2007	2008	2009	2010	Avg. ¹
Jan	152	227	150	113	250	22.0	21.7	22.1	22.6	22.3
Feb	161	135	161	122	193	21.5	21.2	22.2	23.1	22.4
Mar	11	107	64	271	219	22.6	21.1	22.2	22.0	22.6
Apr	7	92	220	68	93	22.2	21.6	21.1	22.2	22.2
May	1	0	77	3	25	21.3	20.1	20.3	21.6	21.0
Jun	0	0	4	0	5	20.1	19.5	19.3	20.4	19.9
Jul	0	0	0	0	5	20.5	19.1	20.2	20.0	19.8
Aug	0	0	52	0	16	20.8	21.4	20.9	21.3	21.6
Sep	0	44	65	0	41	23.6	23.4	23.0	23.3	23.1
Oct	57	17	126	122	129	24.2	24.5	21.9	23.3	23.2
Nov	79	155	113	252	187	22.5	22.0	22.5	21.1	22.5
Dec	298	152	158	309	228	21.8	21.3	21.4	21.7	22.3
Total	766	929	1,190	1,260	1,391					

¹Historical means (1974–2006).

Table 2. Identification, register and species of 10 *Arachis* spp. genotypes.

Identification	BRA germplasm register ¹	Scientific name
Belmonte	031828 (cv. Belmonte)	<i>Arachis pintoii</i>
Mandobi	040550 (cv. BRS Mandobi)	<i>Arachis pintoii</i>
Accession 1	033260	<i>Arachis repens</i>
Accession 2	039799	<i>Arachis pintoii</i>
Accession 3	013251(cv. Amarillo) × 029220	hybrid <i>A. pintoii</i> × <i>A. repens</i>
Accession 4	039187	<i>Arachis pintoii</i>
Accession 5	014991	<i>Arachis pintoii</i>
Accession 6	035114	<i>Arachis pintoii</i>
Accession 7	034436	<i>Arachis repens</i>
Accession 8	034142	<i>Arachis pintoii</i>

¹Official germplasm accession number at Embrapa Recursos Genéticos e Biotecnologia.

Experimental design and sampling

The experimental design was a randomized complete block with 4 replications. Plots consisted of 4 rows of 2 m length with 0.5 m spacing between rows and 0.25 m between plants within rows, totaling an area of 4.0 m². Vegetative material was struck in a greenhouse, using 200-mL plastic cups, and rooted plants were transplanted to plots on 4 December 2007. Plots were harvested on 31 March and 30 May 2008 to estimate dry matter yield (DMY) in the establishment year. Plants were cut at 0.05 m from the soil surface and forage mass was quantified in a 1-m² quadrat placed in the center of the plot. In subsequent years, plots were harvested every 42 days during the rainy season (November–April). The evaluation extended from Dec 2007 to Apr 2010 and harvests were categorized in 3 periods as described earlier

and referred to herein as Year 1, Year 2 and Year 3. Harvests were made as follows: Year 1 - 31 March (1) and 30 May 2008 (2); Year 2 - 4 Dec 2008 (1), 15 Jan (2), 26 Feb (3), 9 Apr (4) and 21 May 2009 (5); and Year 3 - 20 Oct (1), 1 Dec 2009 (2), 12 Jan (3), 23 Feb (4) and 6 Apr 2010 (5).

A few days before each harvest (except for harvest 4 of Year 3), ground cover (GC) percentage of the *Arachis* spp. genotypes was estimated. This visual evaluation was performed by 2 observers who attributed scores to the plots in a range of 0–100%, and these scores were meaned.

Forage samples were dried in a forced-air oven at 55 °C for 72 h and then ground in a Wiley mill fitted with a sieve of 1 mm mesh for subsequent chemical analysis. Dry matter content, and crude protein (CP), neutral detergent fiber (NDF) and acid detergent fiber (ADF)

concentrations were evaluated as described by Silva & Queiroz (2002). In vitro dry matter digestibility (DMD) was determined by the 2-stage method of Tilley & Terry (1963). All samples were analyzed, except those for harvest 1 of Year 3. The concentrations of macronutrients (Ca, Mg, P, K and S) of aerial plant parts were also determined according to Adler and Wilcox (1985). Forage samples from harvest 5 of Year 2 and from harvests 1 and 2 of Year 3 were not analyzed for macronutrient concentrations.

Statistical analyses

Dry matter yield data were analyzed in terms of annual forage accumulation considering all harvests during 3 years of evaluation. Nutritive value variables like DMD and CP, NDF, ADF and macronutrient concentrations were evaluated as annual mean values of forage samples analyzed. Agronomic and chemical analysis data were analyzed using Proc Mixed of SAS (Statistical Analysis System, version 9.2) with genotype, year and genotype \times year as fixed effects, while block was considered a random effect. Year was tested as a fixed effect and a repeated measure because an effect on forage yield in a perennial experiment was expected. However, since genotype \times year interactions were significant for most data ($P < 0.05$), these data were analyzed by year using Proc GLM of SAS. Means for the effect of genotype were

assessed by Least Significant Difference (LSD) by t test ($P < 0.05$). Ground cover (GC) percentage for each harvest was described graphically with standard deviation bars.

Results

Dry matter yield (DMY) was affected by a genotype \times year interaction (Table 3; $P < 0.0001$). In the establishment year, DMY was lower than in Years 2 and 3 (mean yields of 4.2 vs. 8.4 and 8.7 t/ha, respectively). *Arachis pintoi* accessions 2 and 4 were quite productive throughout the evaluation (mean yields of 8.5 and 8.8 t DM/ha, respectively) along with cv. Belmonte (8.8 t DM/ha). Cultivar BRS Mandobi (5.7 t DM/ha) and *A. pintoi* accession 6 (5.6 t DM/ha) were the least productive genotypes throughout the study.

Ground cover was affected by a genotype \times year interaction (Table 3; $P < 0.0001$). In the establishment year GC of the various genotypes ranged from 83 to 96%. In the second and third years, larger differences emerged with ranges of 62–100% in Year 2 and 54–99% in Year 3. Throughout the evaluation, GC was greatest for *A. pintoi* accessions 2 and 4 and cv. Belmonte (Figure 1). Ground cover was maintained above 86% throughout by all genotypes, except for Mandobi and *A. pintoi* accessions 6 and 8, where cover had declined to about 60% or less by the end of the third year.

Table 3. Dry matter yield (DMY) and ground cover (GC) of 10 *Arachis* spp. genotypes in 3 years of evaluation (2008–2010) in Planaltina, Federal District, Brazil.

Genotype ¹	DMY (t/ha)			GC ² (%)		
	Year 1	Year 2	Year 3	Year 1	Year 2	Year 3
Belmonte	4.6	10.5	11.3	96.0	99.7	95.7
Mandobi	3.8	6.4	6.8	88.6	62.1	60.7
Accession 1	2.6	8.8	7.4	92.8	90.1	88.8
Accession 2	5.5	9.6	10.3	87.5	99.8	96.6
Accession 3	3.2	7.2	9.1	95.0	87.5	86.6
Accession 4	5.0	10.4	11.1	92.4	99.5	99.4
Accession 5	4.6	7.8	9.1	91.5	86.0	86.6
Accession 6	3.6	7.0	6.2	82.5	65.1	53.8
Accession 7	4.4	8.4	8.8	89.8	89.8	86.6
Accession 8	4.9	8.2	6.7	91.8	80.0	62.5
LSD ($P < 0.05$)	0.6	2.0	1.5	12.8	13.0	13.1

¹Values presented are averages across 4 replications, with 2 harvests in Year 1 (2008), 5 harvests in Year 2 (2008/09) and 5 harvests in Year 3 (2009/10).

²Ground cover (%) was transformed to (GC/100) arcsin to perform statistical analyses.

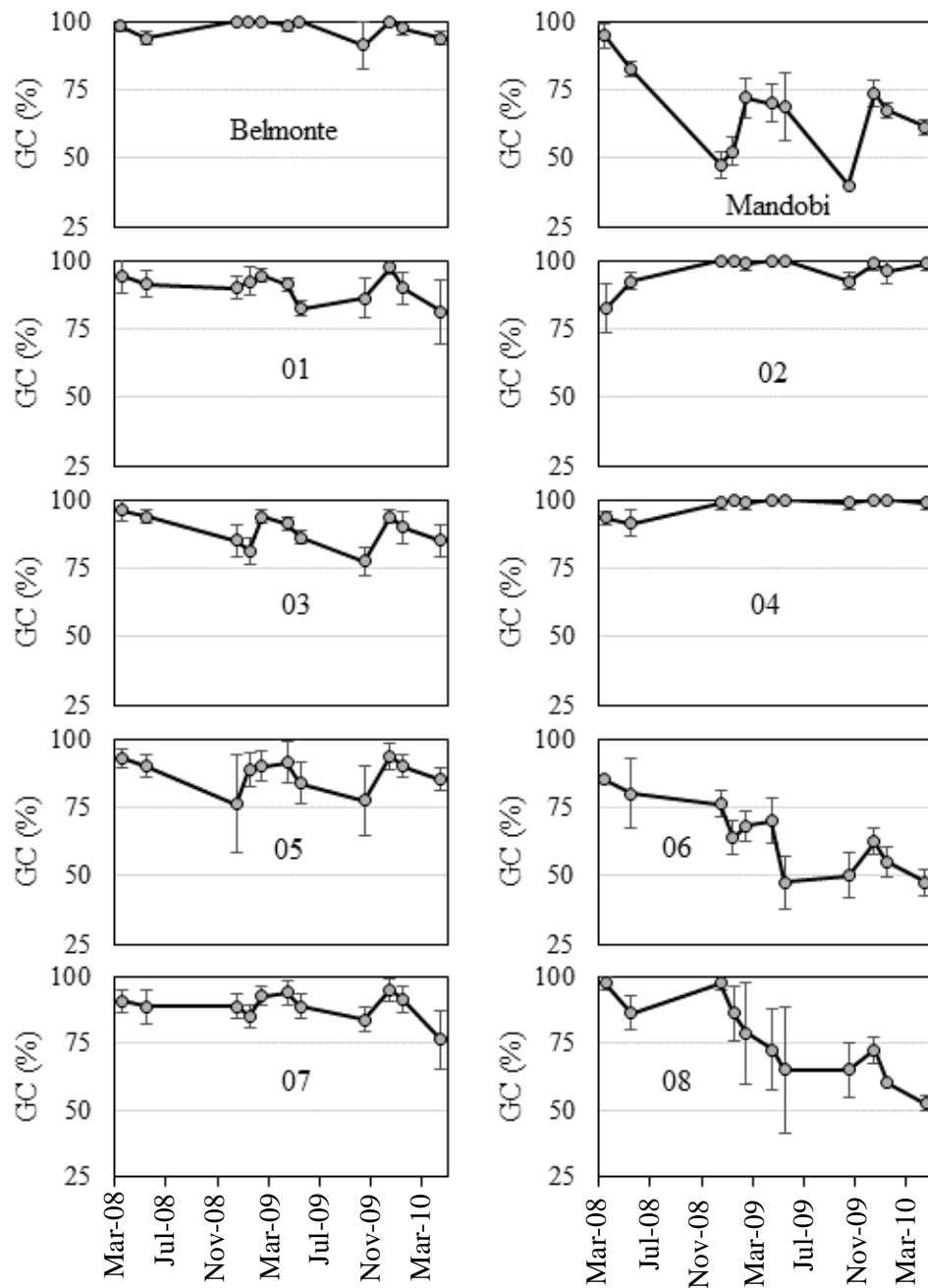


Figure 1. Ground cover (GC) percentage for 10 *Arachis* spp. genotypes in 3 years of evaluation (2008–2010) in Planaltina, Federal District, Brazil. Numbers 1 and 7 represent accessions of *A. repens*, numbers 2, 4, 5, 6 and 8 represent accessions of *A. pintoi* and number 3 represents an *A. pintoi* × *A. repens* hybrid. Small vertical bars represent 2 standard deviations. Each point plotted represents the mean of 4 replicates.

Dry matter digestibility (DMD) was affected by a genotype × year interaction (Table 4; $P = 0.0118$). Mean values were greater for the first 2 years of evaluation (610 and 614 g/kg, respectively) than for the third year (532 g/kg). In the first year, DMD of the hybrid accession 3 (678 g/kg) was 11% superior ($P < 0.05$) to the mean of the remaining genotypes (610 g/kg), but there were no differences between genotypes in the second and third years ($P > 0.05$).

There were significant genotype × year interactions for ADF (Table 4; $P < 0.0001$) and NDF (Table 4; $P = 0.0022$) concentrations. The only consistency in ADF or NDF concentrations in the various genotypes throughout the study was that the hybrid (accession 3) had the lowest ADF values throughout.

Crude protein concentrations were affected by genotype ($P = 0.0004$) and year ($P < 0.0001$). *Arachis repens* accession 7 and *A. pintoi* accessions 2 and 4

Table 4. In vitro dry matter digestibility (DMD), and acid detergent fiber (ADF), neutral detergent fiber (NDF) and crude protein (CP) concentrations of 10 *Arachis* spp. genotypes in 3 years of evaluation (2008–2010) in Planaltina, Federal District, Brazil.

Genotype ¹ , year	DMD (g/kg)			ADF (g/kg)			NDF (g/kg)			CP (g/kg)
	Yr 1	Yr 2	Yr 3	Yr 1	Yr 2	Yr 3	Yr 1	Yr 2	Yr 3	3 years
Belmonte	596	624	530	255	234	259	442	396	408	174
Mandobi	602	615	520	221	233	272	409	400	423	158
Accession 1	602	583	559	241	234	246	462	423	432	162
Accession 2	583	621	524	252	242	258	456	392	403	177
Accession 3	678	618	556	197	210	228	409	386	398	154
Accession 4	602	631	541	232	229	248	425	391	394	176
Accession 5	618	608	501	239	248	280	434	410	432	160
Accession 6	608	599	515	217	233	278	407	403	422	154
Accession 7	597	610	518	252	250	268	445	420	440	182
Accession 8	610	632	556	208	228	249	394	391	400	163
LSD (P<0.05)	38.2	33.9	43.3	10.7	15.7	22.6	26.2	21.5	24.3	13.1
Year 1										142
Year 2										197
Year 3										159
LSD (P<0.05)										5.9

¹Values presented are averages across 4 replications, 2 harvests in Year 1 (2008), 5 harvests in Year 2 (2008/09) and 4 harvests in Year 3 (2009/10).

as well as cv. Belmonte had higher CP concentrations than all other genotypes (Table 4; $P = 0.0004$). In the first year of evaluation, mean CP concentration was 142 g/kg, increasing to 197 g/kg in Year 2, before decreasing to 159 g/kg in Year 3 ($P < 0.0001$).

Concentrations of all macronutrients were affected by genotype \times year interactions ($P < 0.05$) (Table 5). However,

the main difference that occurred in P concentrations was a marked drop from 2.1 and 2.3 g/kg in Years 1 and 2, respectively, to 1.3 g/kg in Year 3. A similar result was found for K concentration, where values dropped from 9.8 and 10.0 g/kg in Years 1 and 2 to 8.3 g/kg in Year 3. Magnesium values also declined from 4.7 and 3.9 g/kg in Years 1 and 2 to 1.5 g/kg in Year 3.

Table 5. Concentrations of macroelements (g/kg DM) in 10 *Arachis* spp. genotypes over 3 years of evaluation (2008–2010) in Planaltina, Federal District, Brazil.

Genotype ¹	P			K			Ca			Mg			S		
	Yr 1	Yr 2	Yr 3	Yr 1	Yr 2	Yr 3	Yr 1	Yr 2	Yr 3	Yr 1	Yr 2	Yr 3	Yr 1	Yr 2	Yr 3
Belmonte	1.8	2.2	1.3	9.0	9.4	5.7	17.5	11.6	22.1	5.0	4.8	2.0	1.6	1.2	1.7
Mandobi	2.1	2.3	1.3	10.9	10.4	10.8	14.9	10.7	19.5	3.5	3.7	1.2	1.8	1.4	1.8
Accession 1	2.7	2.3	1.4	11.0	9.6	7.8	12.0	9.5	18.2	2.8	3.4	1.2	1.5	1.3	1.6
Accession 2	2.0	2.1	1.3	11.1	9.0	6.2	19.1	11.6	23.3	5.1	4.9	2.2	1.8	1.2	1.7
Accession 3	2.1	2.4	1.2	10.3	11.7	8.1	14.8	10.0	16.9	3.8	3.0	0.8	1.5	1.4	1.5
Accession 4	2.1	2.2	1.3	9.4	9.1	6.8	21.5	11.7	24.7	5.3	4.8	2.2	1.7	1.3	1.8
Accession 5	1.9	2.3	1.2	9.4	10.8	8.3	16.1	9.9	14.9	4.7	4.0	1.3	1.5	1.3	1.4
Accession 6	2.1	2.3	1.3	8.8	10.7	12.0	14.8	8.8	14.5	5.0	3.0	1.0	1.6	1.5	1.9
Accession 7	1.8	2.4	1.5	8.4	9.8	7.9	18.5	10.3	18.2	5.5	4.0	1.8	1.5	1.4	1.7
Accession 8	2.2	2.2	1.3	9.2	9.0	9.5	18.2	10.5	15.8	5.9	3.6	1.2	1.8	1.3	1.5
LSD (P<0.05)	0.5	0.2	0.1	2.3	1.3	1.5	3.0	1.1	1.6	1.0	0.7	0.3	0.2	0.2	0.1

¹Values presented are averages across 4 replications, 2 harvests in Year 1 (2008), 4 harvests in Year 2 (2008/09) and 3 harvests in Year 3 (2009/10).

Discussion

Although *Arachis* spp. yields were lower than those of introduced African C4 grasses in Brazil, they were similar to those of native perennial legumes from Brazilian savannas like *Stylosanthes* spp. (Ramos et al. 2010). In Acre State, cv. BRS Mandobi frequently reaches yields of 9–15 t DM/ha in the establishment year (Assis et al. 2013), much higher than the yields obtained in this study (Table 3). Even in Year 3, Mandobi failed to reach yields of that magnitude. On the other hand, cv. Belmonte performed at a higher level and appears better adapted to this environment than Mandobi (Ramos et al. 2010). Assis et al. (2008), in comparing *Arachis* spp. selected for the Amazonian region, observed that *A. pintoii* accessions 2 and 4 presented higher DMVs than other genotypes, including *A. repens*. In this same study, in the establishment year, accessions 2 and 4 yielded about 4 t DM/ha of forage while *A. repens* accessions 1 and 7 yielded about 3 t DM/ha. In Florida, USA, a collection of *A. pintoii* genotypes yielded 4.4 t DM/ha of forage in the first year of evaluation (Carvalho and Quesenberry 2012), similar to the DMVs reported in the first year of the current study. Genotypes of *A. repens* (accessions 1 and 7) showed lower DMVs than *A. pintoii*, which was to be expected for a Brazilian savanna environment (Pizarro and Rincón 1994). Considering the low productivity of most unfertilized grass pastures in Brazilian savannas, the mean DMVs displayed by the *Arachis* spp. could be considered satisfactory, particularly in the second and third years of evaluation.

Persistence of forage peanut in pastures is related to its capacity to cover the ground and thereby prevent the ingress of weeds that leads to pasture degradation. As a strategy to colonize an area, species from the botanical *Arachis* section *Caulorrhizae*, such as *A. pintoii* and *A. repens*, spread by stolons. This clonal reproduction is one of the most efficient mechanisms to ensure the persistence of forage legumes. In addition, the persistence of forage peanut is due to its high tolerance to trampling and defoliation, since it has a prostrate habit, with abundant and well protected growth points. However, the persistence and vigor of *Arachis* spp. in savannas can be short-lived due to irregular distribution of rainfall throughout the year, resulting in severe moisture stress. In 2008, rainfall during the first dry season (May–October) was very low (Table 1) resulting in decline in ground cover of most genotypes, especially Mandobi and accessions 5 and 6. Better rainfall distribution throughout 2009 failed to maintain maximum ground cover for most *Arachis* spp. genotypes during the dry season (Figure 1), because of low minimum temperatures in this period

(<15 °C). However, the ability of these genotypes to maintain such high ground cover until the end of the study demonstrates their ability to survive in edaphoclimatic conditions of savannas. Nevertheless, their performance under grazing, when extra stresses would be experienced, still needs to be evaluated. The genetic variability existing among the germplasm evaluated would enable selection of more adapted genotypes and further gains through breeding. In Acre State, Valentim et al. (2003) observed that Belmonte reached 96% ground cover just 70 days after planting, higher than cv. Amarillo and other genotypes of *A. pintoii* and *A. repens*. For Amazonian conditions, Assis et al. (2008) observed that cvv. Belmonte and BRS Mandobi and accessions 2 and 4 were the best performing genotypes during the establishment phase in terms of DM yield and ground cover. Except for Mandobi, these results were similar to our findings under savanna conditions in the current study (Figure 1). In general, the most productive genotypes in the rainy season were those which achieved the highest values for ground cover.

As establishment of *Arachis* spp. is considered slow, the lesser productivity is continually associated with stand formation problems, even in humid areas (Valentim et al. 2003). In the current study, the slow initial development limited the number of cuts (2) in the first rainy season. This can require additional resources to assure an adequate initial ground cover and forage production (Carvalho and Quesenberry 2012), e.g. new germplasm and fertilization strategies. Otherwise, the expected benefits of the legume in the short term can be reduced (Pizarro and Rincón 1994). Because of its ability to produce acceptable seed yields, cv. BRS Mandobi was chosen from a selection network including many regions of Brazil. Assis et al. (2013) stated that about 18–21 months after planting, Mandobi produced 3 t/ha of pure seeds. This is an important characteristic, considering the restrictions on farmer adoption of *Arachis* spp. caused by the need for vegetative propagation, as can be observed for cv. Belmonte, which produces little seed. Although DM yield and persistence in savannas of some *Arachis* spp. might not be satisfactory, particularly for cv. BRS Mandobi, high seed production would allow for a lesser seed price, which could compensate for the need to have to replant the legume periodically. However, as a consequence of the geocarpic reproductive growth, lack of relevant seed harvesting technology could increase costs of seed production (Ferguson 1994).

In general, dry matter digestibility (DMD) of *Arachis* spp. is greater than that of other herbaceous tropical legumes, regardless of the season of the year (Gama et al. 2014). As it is strongly associated with forage quality and

systems during the rainy season, through a direct contribution to animal diets and also indirectly through promoting substantial recycling of nitrogen in the soil in mixed legume-grass pastures and increasing their longevity. While introduction, management and persistence of *Arachis* spp. are still relevant issues, seed production technology is one of the most important aspects that should be addressed in order to provide affordable and readily accessible seed for farmers. Under savanna conditions, in terms of forage production, nutritive value, ground cover and tissue macronutrient concentration, the new *A. pintoii* accessions 2 and 4 (accessions BRA-039799 and BRA-039187, respectively) were considered the most promising in this study. Their performance under grazing needs to be evaluated and steps made to improve seed production of these new materials.

References

- Adjolohoun S; Bindelle J; Adandedjan C; Toleba SS; Houinato M; Sinsin B. 2013. Variety and environmental effects on crude protein concentration and mineral composition of *Arachis pintoii* (Krapovickas & Gregory) in Benin (West Africa). *Journal of Applied Biology and Biotechnology* 1:24–28. DOI: [10.7324/JABB.2013.1304](https://doi.org/10.7324/JABB.2013.1304)
- Adler PR; Wilcox GE. 1985. Rapid perchloric acid digest methods for analysis of major elements in plant tissue. *Communications in Soil Science and Plant Analysis* 16:1153–1163. DOI: [10.1080/00103628509367673](https://doi.org/10.1080/00103628509367673)
- Argel PJ; Pizarro EA. 1992. Germplasm case study: *Arachis pintoii*. In: Hardy B, ed. Pastures for the tropical lowlands - CIAT's contribution. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. p. 57–76. (Available at: <https://goo.gl/mx85wP>).
- Assis GML; Valentim JF; Carneiro Jr. JMC; Azevedo JMA; Ferreira AS. 2008. Seleção de genótipos de amendoim forrageiro para cobertura do solo e produção de biomassa aérea no período de estabelecimento utilizando-se metodologia de modelos mistos. *Revista Brasileira de Zootecnia* 11:1905–1911. DOI: [10.1590/S1516-35982008001100001](https://doi.org/10.1590/S1516-35982008001100001)
- Assis GML; Valentim JF; Andrade CMS. 2013. BRS Mandobi: A new forage peanut cultivar propagated by seeds for the tropics. *Tropical Grasslands-Forrajes Tropicales* 1:39–41. DOI: [10.17138/TGFT\(1\)39-41](https://doi.org/10.17138/TGFT(1)39-41)
- Boddey RM; Sá JCM; Alves BJR; Urquiaga S. 1997. The contribution of biological nitrogen fixation for sustainable agricultural systems in the tropics. *Soil Biology and Biochemistry* 29:787–799. DOI: [10.1016/S0038-0717\(96\)00221-0](https://doi.org/10.1016/S0038-0717(96)00221-0)
- Carvalho MA; Quesenberry KH. 2012. Agronomic evaluation of *Arachis pintoii* (Krap. and Greg.) germplasm in Florida. *Archivos de Zootecnia* 61:19–29. DOI: [10.4321/s0004-05922012000100003](https://doi.org/10.4321/s0004-05922012000100003)
- Ferguson JE. 1994. Seed biology and seed systems for *Arachis pintoii*. In: Kerridge PC; Hardy B, eds. Biology and agronomy of forage *Arachis*. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. p. 122–133. (Available at: <https://goo.gl/UlyiEq>).
- Gama TCM; Volpe E; Lempp B. 2014. Biomass accumulation and chemical composition of Massai grass intercropped with forage legumes on an integrated crop-livestock-forest system. *Revista Brasileira de Zootecnia* 43:279–288. DOI: [10.1590/S1516-35982014000600001](https://doi.org/10.1590/S1516-35982014000600001)
- Lascano CE. 1994. Nutritive value and animal production of *Arachis*. In: Kerridge PC; Hardy B, eds. Biology and agronomy of forage *Arachis*. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. p. 109–121. (Available at: <https://goo.gl/UlyiEq>).
- Oliveira OC; Oliveira IP; Alves BJR; Urquiaga S; Boddey RM. 2004. Chemical and biological indicators of decline/degradation of *Brachiaria* pastures in the Brazilian Cerrado. *Agriculture, Ecosystems and Environment* 103:289–300. DOI: [10.1016/j.agee.2003.12.004](https://doi.org/10.1016/j.agee.2003.12.004)
- Paganella MB; Valls JFM. 2002. Caracterização morfológica de cultivares a acessos selecionados de *Arachis pintoii* Krapov. & Gregory. *Pasturas Tropicales* 24(2):23–30. (Available at: <https://goo.gl/spLKRh>).
- Peel MC; Finlayson BL; McMahon TA. 2007. Updated world map of the Köppen-Geiger climate classification. *Hydrology Earth System Sciences* 11:1633–1644. DOI: [10.5194/hess-11-1633-2007](https://doi.org/10.5194/hess-11-1633-2007)
- Pizarro EA; Rincón A. 1994. Regional experience with forage *Arachis* in South America. In: Kerridge PC; Hardy B, eds. Biology and agronomy of forage *Arachis*. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. p. 144–157. (Available at: <https://goo.gl/UlyiEq>).
- Rao IM; Kerridge PC. 1994. Mineral nutrition of forage *Arachis*. In: Kerridge PC; Hardy B, eds. Biology and agronomy of forage *Arachis*. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. p. 71–83. (Available at: <https://goo.gl/UlyiEq>).
- Ramos AKB; Barcellos AO; Fernandes FD. 2010. Gênero *Arachis*. In: Fonseca DM; Martuscello JA, eds. Plantas forrageiras. Editora da UFV, Universidade Federal de Viçosa, Viçosa, MG, Brazil. p. 249–293.
- Silva DJ; Queiroz AC. 2002. Análise de alimentos: Métodos químicos e biológicos. 3rd Edn. Universidade Federal de Viçosa (UFV), Viçosa, MG, Brazil.
- Tilley JMA; Terry RA. 1963. A two-stage technique for the *in vitro* digestion of forage crops. *Grass and Forage Science* 18:104–111. DOI: [10.1111/j.1365-2494.1963.tb00335.x](https://doi.org/10.1111/j.1365-2494.1963.tb00335.x)
- Valentim JF; Andrade CMS; Mendonça HA; Sales MFL. 2003. Velocidade de estabelecimento de acessos de amendoim forrageiro na Amazônia Ocidental. *Revista Brasileira de Zootecnia* 32:1569–1577. DOI: [10.1590/S1516-35982003000700005](https://doi.org/10.1590/S1516-35982003000700005)

Valls JFM; Simpson CE. 1994. Taxonomy, natural distribution, and attributes of *Arachis*. In: Kerridge PC; Hardy B, eds. Biology and agronomy of forage *Arachis*. Centro Internacional de Agricultura Tropical (CIAT), Cali, Colombia. p. 1–18. (Available at: <https://goo.gl/UlyiEq>).

Wünscher T; Schultze-Kraft R; Peters M; Rivas L. 2004. Early adoption of the tropical forage legume *Arachis pinto* in Huetar Norte, Costa Rica. Experimental Agriculture 40:257–268. DOI: [10.1017/S0014479703001583](https://doi.org/10.1017/S0014479703001583)

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